This article was downloaded by: On: 29 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37- 41 Mortimer Street, London W1T 3JH, UK

Supramolecular Chemistry

Publication details, including instructions for authors and subscription information: <http://www.informaworld.com/smpp/title~content=t713649759>

Macrocyclic polyethers incorporating resorcinol residues as templates for cyclobis(paraquat-p-phenylene) in the self-assembly of [2]catenanes

David B. Amabilino^a; Peter R. Ashton^a; J. Fraser Stoddart^a a School of Chemistry, The University of Birmingham, Edgbaston, Birmingham, UK

To cite this Article Amabilino, David B. , Ashton, Peter R. and Stoddart, J. Fraser(1995) 'Macrocyclic polyethers incorporating resorcinol residues as templates for cyclobis(paraquat-p-phenylene) in the self-assembly of [2]catenanes', Supramolecular Chemistry, 5: $1, 5 - 8$

To link to this Article: DOI: 10.1080/10610279508029880 URL: <http://dx.doi.org/10.1080/10610279508029880>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use:<http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

COMMUNICATION

Macrocyclic polyethers incorporating resorcinol residues as templates for cyclobis(paraquat-p-phenylene) in the selfassembly of [2]catenanes[†]

DAVID B. AMABILINO, PETER R. ASHTON and J. FRASER STODDART*

School of Chemistry, The University of Birmingham, Edgbaston, Birmingham Bl5 2TI: UK

+This communication is dedicated to Professor Donald J. Cram on the occasion of his 75th birthday.

{Received **March** *11,1994)*

The self-assembly of [2]catenanes incorporating cyclobis(paraquat-p-phenylene) and either bis(metaphenylene)-32 crown-10 or tris(metaphenylene)-48-crown-l5 has been achieved. The dynamic processes associated with the relative motions of the **two rings have been studied by variable temperature 'H NMR spectroscopy. Both [2]catenanes display rapid relative movements of the two components, associated with free energies of activation in the region 12-14 kcal m0l-l for different processes. The resorcino1 residues of the macrocyclic polyethers** *are* **bound in the cavity of the cyclophane by** *m-m* **stacking, electrostatic interactions, and T-type hydrogen bonding.**

An understanding of the relationship between the strength and geometry of binding in host-guest $1-3$ complexes, and the ability of such supramolecular systems to undergo self-assembly4-5 relies, at present, upon **an** empirical structural and synthetic chemical foundation.6 The formation of catenanes⁷ requires initially the complexation of a linear component threaded through the center of a macrocyclic component. Cyclisation of the linear component then affords a [2]catenane. The solid state 1:1 complex $8-9$ formed between bis(paraphenylene)-34-crown- 10 (BPP34C10) and Paraquat paved the way^{7,10} for the template-directed synthesis of $[2]$ - and [3]-catenanes comprised of these building blocks. There is, however, no solid-state structural evidence^{11} concerning the geometry of the binding of Paraquat by the crown ether bis(metaphenylene)-32-crown-10 (BMP32C10). One extremely appealing way **to** test binding geometry is simply to attempt the self-assembly of a [2]catenane incorporating **cyclobis(paraquat-p-phenylene).** Here, we describe the successful self-assembly and the characterization of two new [2]catenanes in which the macrocyclic polyether components contain resorcinol residues.

The macrocyclic polyethers BMP32C10 and **tris(metaphenylene)-48-crown-15** (TMP48C15)12 were prepared (Scheme 1) in 8% and 2% yields, respectively, in a single step (Scheme 1) by the cesium carbonate-promoted reaction¹³ of resorcinol $(1,3$ -dihydroxybenzene) with the bistosylate of tetraethylene glycol. The major product of this reaction, in **54%** yield, is the crown ether¹⁴ metaphenylene-16-crown-5 (MP16C5). The isolated macrocyclic polyethers BMP32C10 and TMP48C15, which were separated by chromatography, were each reacted under identical conditions (RT) with l.2PF6 and **1,4-bis(bromornethyl)benzene** in DMF. After one week, the reaction mixtures were subjected to column chromatography, affording (Scheme 2) the [2]catenanes 2.4PF₆ and 3.4PF₆ as yellow solids¹⁵⁻¹⁶ in 17% and 12% yields, respectively. They were characterized by variable temperature ¹H NMR spectroscopy and fast atom bombardment mass spectrometry (FABMS).

The FABMS of both catenanes show the characteristic sequential loss of PF_6 counterions, as well as decatenation in the spectrometer. The highest mass peak for

^{*}To whom correspondence should **be.** addressed.

Scheme 1 The synthesis of the macrocyclic polyethers MP16C5, BMP32C10, and TMP48C15 in one step. (See Color Plate I.)

2.4PF₆ was observed at m/z 1490, corresponding to [M- PF_6 ⁺, while 3.4PF₆ gave a peak with m/z 1928, which corresponds to the [M+Na]+ ion. Since the [2]catenane $3.4PF₆$ has polyether chains which are not involved in noncovalent bonding interactions with the cyclophane, it presumably binds more readily sodium ions in the metanitrobenzyl alcohol matrix.

The ¹H NMR spectra (400 MHz) of both $2.4PF₆$ and $3.4PF₆$ at ambient temperature reveal well-resolved resonances arising from the cyclophane components of the structures, while the resonances for **the** protons attached to the resorcinol residues are broadened into the baseline of the spectrum on account of an exchange process in which the macrocyclic polyether circumrotates through the cavity of the tetracationic cyclophane (Process I in Figure 1).

When a CD_3COCD_3 solution of 2.4PF₆ is cooled down to O"C, the signals, which correspond to the resorcinol residues of the BMP32C10 component residing 'inside' and 'outside' the tetracationic component of the [2]catenane, become evident in the IH NMR spectrum. The free energy barrier to the hindered circumrotation of the macrocyclic polyether component through the cavity of the tetracationic cyclophane (Process I in Figure 1) was calculated to be $\Delta G_c^{\dagger} = 13.6$ kcal mol⁻¹ based on the coalescence17 at 304 K of the H-416 protons attached to the

'inside' and 'outside' resorcinol rings $(\Delta v = 513 \text{ Hz}$ and $k_c = 1160 \text{ s}^{-1}$). In the case of 2.4PF₆, Process II (Figure 1) remains fast on the ¹H NMR timescale down to 193 Kbelow which temperature CD_3COCD_3 freezes. However, an additional process with a free energy barrier $\Delta G_c^{\dagger} =$ 12.1 kcal mol-1 (average value) was calculated from the coalescences at 251 K and 240 K, respectively, of the CH₂ ($\Delta v = 58$ Hz and $k_c = 129$ s⁻¹) and α -CH ($\Delta v = 27$ Hz and $k_e = 60$ s⁻¹) proton resonances arising from the tetracationic cyclophane. This process corresponds to the energy required to equilibrate (Process 111 in Figure 2) the two possible orientations¹⁸ of the resorcinol residue within the π -electron deficient cavity. A resorcinol ring leaves the cavity of the cyclophane and is replaced with one having an orientation where the H-2 **and** H-5 protons have switched their positions relative to those occupied in the initial structure. The free energy barrier to the analogous process in $3.4PF_6$ in CD_3COCD_3 solution has a similar value of 12.2 kcal mol⁻¹ for its ΔG_c^{\dagger} , calculated from the coalescence temperature of 248 K ($\Delta v = 38$ Hz and $k_c = 85 \text{ s}^{-1}$) of the α -CH protons of the cyclophane.¹⁹

As the solution of $2.4PF_6$ is cooled down, the resonances arising from the protons attached to both the 'inside' and 'outside' resorcinol units shift significantly. In particular, the protons at the 2- and 5-positions of the included ring exhibit resonances which shift (Figure 3)

Scheme 2 The template-directed self-assembly of the [2]catenanes $2.4PF_6$ and $3.4PF_6$. (See Color Plate II.)

Figure 1 A diagrammatic representation of the two of the dynamic processes (I and 11) taking place between the two components of the [2]catenane 2.4PF,. (See Color Plate 111.)

from 6 1.73 and **3.06** at 253 **K,** to **6 1.99** and 2.3 1, respectively, at 193 K. Meanwhile, the resonances of the **H-5** and the equivalent H-4 and **H-6** protons attached to the 'outside' resorcinol unit shift from 6 6.71 and **6.15** at 253 **K,** to 6 6.23 and 5.91, respectively, at 213 K, and the chemical shift of the resonance arising from the H-2 proton is essentially invariant with temperature. Interestingly, chemical shift changes with temperature observed for H-2 and H-5 in $3.4PF_6$ are not as dramatic as those observed for the same protons in $2.4PF_6$. The H-2 proton resonance shifts from δ 1.70 at 253K to δ 1.80 at 213 K, while the H-5 proton was revealed at δ 3.85 by

Figure 2 A diagrammatic representation of the equilibration (Process 111) of **the two possible degenerate orientations** of **resorcinol units within the cavity** of **cyclobis(paraquat-p-phenylene) in the [2]catenanes. (See Color Plate IV.)**

Figure 3 The temperature dependence of the resonances for H-2 and H-5 arising from the 'inside' resorcinol rings in the 'H NMR spectrum of 2.4PF,. (See Color Plate V.)

a saturation transfer experiment performed at 265 K. The exact reasons for these chemical shift changes are currently being investigated.

The differences in the free energies of activation for the dynamic processes associated with the relative motions of the components in the two [2]catenanes described here, compared with those observed in isomeric [2]catenanes, $7,19$ indicates that the interaction of resorcinol rings in the macrocyclic polyether component with the bipyridinium units in **cyclobis(paraquat-p-phenylene)** is not as significant as the interactions when hydroquinone rings replace the resorcinol rings in the macrocyclic polyether component. In particular, for the isomeric [2]catenane incorporating **BPP34C10**, the free energy barrier⁷ to Process I (Figure 1) is 15.6 kcal mol⁻¹, and that to Process II is 12.2 kcal mol⁻¹. In $2.4PF_6$, the activation barrier to Process I is only **13.6** kcal mol-l, and Process **I1** is clearly also associated with a much lower free energy barrier. The relatively poor yields obtained in the catenations also reflect the detrimental influence that constitutional change²⁰ has upon host-guest interactions¹⁻³ and associated molecular self-assembly processes. The yield of 2.4PF₆ is 17%, while the isomeric [2]catenane containing the macrocyclic ether BPP34C10 was isolated⁷ in 70% yield. A detailed understanding of the structural factors that affect the efficiency of self-assembly processes²¹ is essential if this approach to synthesis is going to lead to the rapid and precise creation of nanometer scale molecular and supramolecular structures.

ACKNOWLEDGEMENTS

We thank the Science and Engineering Research Council in the UK for the financial support of this research.

REFERENCES

- 1 Cram, D.J.; Cram, J.M.; *Acc.* Chem. *Res.* 1978, *11,* 8.
- 2 Cram, D.J.; Trueblood, K.N.; Top. Curr. Chem. 1981, 98, 43.
- 3 Cram, D.J.; *Angew.* Chem. *Inr. Ed. Engl.* 1988,27, 1009.
- 4 Lindsey, J.S.; *New* J. Chern. 1991, *IS,* 153.
- **5** Philp, D.; Stoddart, J.F.; *Synlerf* 1991,445.
- 6 Gokel, G.W.; Medina, J.C.; Li, C.; *Synlert* 1991,677
- 7 Anelli, P.L.; Ashton, P.R.; Ballardini, R.; Balzani, V.; Delgado, M.; Gandolfi, M.T.; Goodnow. T.T.; Kaifer. A.E.; Philp. D.; Pietraszkiewicz, M.; Prodi, L.; Reddington, M.V.; Slawin, A.M.Z.; Spencer, N.; Stoddart, J.F.: Vicent, C.; Williams, D.J.; J. *Am.* Chem. *Soc.* 1992,114, 193.
- 8 Allwood, B.L.; Spencer, N.; Shahriari-Zavareh, H.; Stoddart, J.F.; Williams, D.J.; J. Chem. *Soc.,* Chem. *Commun.* 1987, 1064.
- 9 Helgeson, R.C.; Tamowski, T.L.; Timko, J.M.; Cram, D.J.; J. *Am.* Chem. *Sac.* 1977,99,6411.
- 10 Ashton, P.R.; Brown, C.L.; Chrystal, E.J.T.; Parry, K.P.: Pietraszkiewicz, M.; Spencer, N.; Stoddart, J.F.; Angew. Chem. Int. *Ed. Engl.* 1991.30, 1042.
- 1 **I** Allwood, B.L.; Shahriari-Zavareh, H.; Stoddart. J.F.; Williams, D.J.; J. Chem. *Soc., Chem. Commun.* 1987, 1058.
- 12 MP16C15 had m/z (positive-ion EIMS) 268 corresponding to $[M]$ ⁺; ¹H NMR (CDCl₃, 300 MHz) 3.52-3.59 (4H, m), 3.60-3.65 (4H, m), 3.74 (4H, t). 4.24 (4H, t), 6.47-6.54 (2H. m), 7.04-7.12 (2H, m); ¹³C NMR (CDCl₃, 75 MHz) 68.8, 70.4, 70.7, 70.8, 103.8, **110.1,** 129.4, 160.3.
- 13 Ostrowicki, A.; Koepp, *E.;* Vogtle, F.; *Top. Curr* Chem. 1991, 161,37.
- 14 The analytical data for BMP32C10 has been reported previously (ref. 11). **TMP48C15** had m/z (positive-ion FABMS) 805, corresponding to $[M+H]^+$; ¹H NMR (CDCl₃, 300 MHz) 3.64-3.74 $(24H, m)$, 3.82 (12H, t), 4.08 (12H, t), 6.47–6.52 (9H, m), 7.09-7.18 (3H, m); ¹³C NMR (CDCl₃, 75 MHz) 67.5, 69.7, 70.7, 70.9, 101.9, 107.2, 129.8, 160.0.
- 15 $2.4PF_6$ had m/z (positive-ion FABMS) 1490, 1345 and 1200, corresponding to, $[M-PF_6]^+$, $[M-2PF_6]^+$, and $[M-3PF_6]^+$; ¹H NMR

(CD3COCD3, 400 MHz, 253K) 1.73 **(IH,** bt), 3.05 (IH, t), 3.45-3.55 (4H, **m),** 3.65-3.95 (28H. m). 4.76 (2H, 9). **5.61** (IH, bt), 5.95-6.10 (10H, m), 6.71 (1H, t), 8.04 (8H, bd), 8.25-8.40 $(8H, m)$, 9.38 $(8H, bd)$; ¹³C NMR $(CD_3COCD_3, 101 MHz, 233K)$ 64.9, 65.1, 67.3, 67.4, 69.8 (X 2). 70.1 (X2), 70.5, 71.1. 99.4, 100.5, 105.8, 106.8, 126.1, 127.1, 128.2, 130.9, 131.3, 131.4, 137.9, 138.3, 145.1, 145.6, 146.8, 146.9, 157.7, 160.0.

- 16 3.4PF₆ had *m/z* (positive-ion FABMS) 1928, 1759, 1614 and 1469, corresponding to $[M+Na]^+$, $[M-PF_6]^+$, $[M-2PF_6]^+$, and $[M-3PF_6]^+$; ¹H NMR (CD₃COCD₃, 400 MHz, 253K) 1.70 (1H, bt), 3.40–3.50 $(4H, m)$, 3.60-4.00 (45H, m), 4.43 (2H, d), 5.90-6.10 (10H, bm), 6.26 (2H, d), 6.30 (2H, d), 7.04 (2H, t), 8.02 **(8H, bd),** 8.10-8.30 (8H, bm), 9.36 (8H, bd); ¹³C NMR (CD₃COCD₃, 101 MHz, 233K) 65.1, 65.2, 67.6 (X2). 69.3, 69.4, 69.5, 69.6 (XZ), 69.8, 70.5 (X3). 70.9, 101.2 (X2). 104.5, 106.3, 106.9, 126.4, 127.0, 130.0, 130.9. 131.3 (X2). 137.7, 138.2, 145.1, 145.6, 146.9, 147.2, 157.7, 160.0. 160.2.
- 17 A value for k_c was obtained (Sutherland, I.O. *Annu. Rep. NMR Spectrosc.* **1971**, 4, 71) by using the approximate expression k_c = $\pi(\Delta v)$ / (2)^{1/2}. Using the Eyring equation, this rate constant was used to calculate the ΔG_c^{\dagger} value at the coalescence temperature.
- 18 A similar process has been observed in a [2]catenane. See Ashton, P.R.; Brown, C.L.; Chrystal, E.J.T.; Goodnow, **T.T.;** Kaifer, A.E.; Parry, K.P.; Philp, D.; Slawin, A.M.Z.; Spencer. N.; Stoddart, J.F.: Williams, D.J.; J. Chem. *Soc.,* Chem. *Commun.* 1991, 634.
- A separation of the H-4/6 proton resonances for 'inside' and 'outside' resorcinol residues in $3.4PF_6$ is observed at 308 K in $CD₃COCD₃$ solution. This temperature dependent process reflects the movement of the tetracationic cyclophane between each of the three resorcinol residues in the **TMP48C15** component. In $3.4PF_6$, the ratio of 'inside' to 'outside' resorcinol residues is 1.2. Since the method described in reference 17 strictly speaking only applies to situations where the exchanging signals have equal intensities, it is not possible to calculate a value of ΔG_c^{\dagger} for this process using this treatment.
- 20 Amabilino, D.B.; Ashton, P.R.; Tolley, M.S.; Stoddart, J.F.; Williams, D.J.; *Angew.* Chem *Inr. Ed. Engl.* 1993, 32, 1297.
- 21 Amabilino, D.B.; Stoddart. J.F.; Pure *Appl.* Chem. 1993,65,2351.